

## 10: ASW:BG

Final volume: 1000 ml

### **Mail sol. C<sub>10</sub>**

<b>ASW (Artificial Seawater)</b>	500.00	ml
<b>BG11 (Blue-Green Medium)</b>	500.00	ml

1. 1:1 mixture
2. See separate recipes ASW and BG11. This medium is made up in 2 parts which are autoclaved separately and mixed aseptically when cool. Note: vitamins are not required in the ASW part of this recipe.

### **ASW (Artificial Seawater)**

Tricine	0.50	g
<b>Extra salts</b>	3.75	ml
<b>SE1 (Soil Extract 1)</b>	25.00	ml

1. Make up to 1 litre with filtered natural sea water. Adjust the pH to 7.6 – 7.8 with 1N NaOH or 1N HCl. Autoclave at 15 psi.
2. Alternatively use "Ultramarine Synthetica" sea salts 33.6 g and make up to 1 litre with distilled water. Adjust pH as above.

### **Extra salts**

NaNO <sub>3</sub>	30.00	g
Na <sub>2</sub> HPO <sub>4</sub>	1.20	g
K <sub>2</sub> HPO <sub>4</sub>	1.00	g

### **SE1 (Soil Extract 1)**

Soil (air-dried)	traces
Distilled water	traces

1. Site selection for a good soil is very important and for most purposes a soil from undisturbed deciduous woodland is best. Sites to avoid are those showing obvious signs of man's activity and particular care should be taken to avoid areas where fertilizers, crop sprays or other toxic chemicals may have been used.
2. A rich loam with good crumb structure should be sought. Stones, roots and larger invertebrates should be removed during an initial sieving through a 1 cm mesh. The sieved soil should be spread to air dry and handpicked for smaller invertebrates and roots. It

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should be turned periodically and picked over again. When dry sieve through a finer mesh (2-4 mm) and store in an airtight container away from light and heat.

3. Soil is prepared as above. Air-dried soil and twice its volume of supernatant distilled water are autoclaved together at 15 psi for 2 hours and left to cool. The supernatant is then decanted and filtered through Whatman No 1 filter paper, then distributed to containers in volumes suitable for making up batches of media. The aliquots and their containers are autoclaved for an appropriate length of time (e.g. 1 litre or less for 15 minutes) and are then kept in a cool place (e.g. a refrigerator) until required.

### BG11 (Blue-Green Medium)

NaNO <sub>3</sub> (15 g/l stock solution)	100.00	ml
K <sub>2</sub> HPO <sub>4</sub> (4 g/l stock solution)	10.00	ml
MgSO <sub>4</sub> x 7 H <sub>2</sub> O (7.5 g/l stock solution)	10.00	ml
CaCl <sub>2</sub> x 2 H <sub>2</sub> O (3.6 g/l stock solution)	10.00	ml
Citric acid (0.6 g/l stock solution)	10.00	ml
Ammonium ferric citrate (green, 0.6 g/l stock solution)	10.00	ml
Na <sub>2</sub> -EDTA (0.1 g/l stock solution)	10.00	ml
Na <sub>2</sub> CO <sub>3</sub> (2.0 g/l stock solution)	10.00	ml
<b>Trace metal solution</b>	1.00	ml
Agar, for solid medium	15.00	g

Make up to 1 litre with deionized water. Adjust pH to 7.1 with 1M NaOH or HCl. For agar add 15.0 g per litre of Bacteriological Agar (Oxoid L11). Autoclave at 15 psi for 15 minutes.

### Trace metal solution

H <sub>3</sub> BO <sub>3</sub>	2.86	g
MnCl <sub>2</sub> x 4 H <sub>2</sub> O	1.81	g
ZnSO <sub>4</sub> x 7 H <sub>2</sub> O	0.22	g
Na <sub>2</sub> MoO <sub>4</sub> x 2 H <sub>2</sub> O	0.39	g
CuSO <sub>4</sub> x 5 H <sub>2</sub> O	0.08	g
Co(NO <sub>3</sub> ) <sub>2</sub> x 6 H <sub>2</sub> O	0.05	g