

666: SUCCINICLASTICUM MEDIUM

This recipe contains strain-specific modifications for *unclassified bacterium* DSM 11001 *

Final pH: 6.7 - 6.8

Final volume: 1000 ml

Clarified rumen fluid	400.00	ml
K ₂ HPO ₄	0.23	g
KH ₂ PO ₄	0.23	g
NaCl	0.45	g
(NH ₄) ₂ SO ₄	0.45	g
CaCl ₂ x 2 H ₂ O	0.06	g
MgSO ₄ x 7 H ₂ O	0.09	g
Indigocarmine	5.00	mg
NaHCO ₃	6.40	g
Disodium succinate	5.00	g
Yeast extract	5.00	g
L-Cysteine HCl x H ₂ O	0.30	g
Na ₂ S x 9 H ₂ O	0.30	g
Trypticase peptone	5.00	g
Na-DL-lactate	5.00	g
Distilled water	600.00	ml

1. Dissolve ingredients (except bicarbonate, succinate, yeast extract, cysteine and sulfide), bring medium to the boil, then cool to room temperature under 100% CO₂ gas atmosphere. Add the bicarbonate and equilibrate the medium with the CO₂ gas to pH 6.8. Distribute under 100% CO₂ gas atmosphere into anoxic Hungate-type tubes or serum vials and autoclave. Thereafter, add succinate, yeast extract, cysteine and sulfide from sterile anoxic stock solutions prepared under 100% N₂ gas atmosphere. Adjust pH of complete medium to 6.7 - 6.8, if necessary.

2. Note: Supplementing the medium with 1.50 g/l agar stimulates growth of strains after resuscitation from ampoules.

* Omit yeast extract and succinate and supplement medium with 5.00 g/l each of Trypticase peptone and Na-(DL)-lactate.

Clarified rumen fluid (from medium 1310)

Rumen fluid from cow or sheep (obtained from fistulated animals or abattoir refuse) is filtered through muslin, autoclaved at 121°C for 15 min and then centrifuged at 27,000 g for 20 min. The supernatant is made anoxic by sparging with 100% N₂ gas for 15 min, dispensed under same gas atmosphere into anoxic serum vials to 30% of volume and then stored frozen at -20°C.